CRANE AND WINTER FLIES (DIPTERA: LIMONIIDAE, PEDICIIDAE, TRICHOCERIDAE) ASSOCIATED WITH FUNGI IN LITHUANIA

Virginija PODĖNIENĖ, Jolanta RIMŠAITĖ, Sigitas PODĖNAS

Department of Zoology, Vilnius University, Čiurlionio 21/27, LT-03101 Vilnius, Lithuania. E-mail: sigitas.podenas@gf.vu.lt Nature Research Centre, Akademijos st. 2, LT-08412 Vilnius, Lithuania. E-mail: jolanta@ekoi.lt Nature Research Centre, Akademijos st. 2, LT-08412 Vilnius, Lithuania. E-mail: sigitas.podenas@ekoi.lt

Abstract. The list of Lithuanian crane flies and winter flies, whose larvae were found developing in fungi, is given. The list of fungi, in which larvae of crane and winter flies develop, is compiled in Lithuania for the first time. For the first time were larvae of *Metalimnobia bifasciata* (Schrank, 1781) reared from seven species of fungi and *Ula bolitophila* Loew, 1869 was first reared from one fungus species. This study revealed one new for Lithuania species of crane flies – *Ula mixta* Stary, 1983. In addition, for the first time a trophic relationship between this species of crane flies and fungi was revealed.

Key words: Limoniinae, Pediciidae, Trichoceridae, larvae, Limonia, Metalimnobia, Ula, Trichocera

Introduction

Larvae of 25 species belonging to two families of Tipuloidea crane flies (Limoniidae and Pediciidae) can develop in various fungus species in the Palaearctic. Larvae of Tipulidae crane flies can be found in such a habitat as fungi only occasionally. Development of some Limoniidae and Pediciidae species can take place only in fungi, whereas larvae of other species can be found both in mouldy wood and mushrooms. Species of crane flies associated with fungi are studied in other countries comparatively well. The first papers devoted to that issue appeared in the 19th century (Stannius 1829). Lindner's (1958) publication appeared to be the first purposive study of crane flies developing in fungi. Other authors (Buxton 1960; Dely-Draskovits 1972; Trifourkis 1977; Hackman & Meinander 1979; Savchenko & Krivolutskaya 1976; Stubbs & Chandler 1978; Krivosheina et al. 1986; Yakovlev 1994; Ševčik 2001, 2006) compiled the list of mushroom species associated with crane fly larvae. Only larvae of the species belonging to the genera Achyrolimonia, Atypophthalmus, Discobola, Limonia, Metalimnobia, Rhipidia (Limoniidae, Limoniinae) and Ula (Pediciidae) develop in fungi. Scleroprocta sororcula (Zetterstedt, 1851), which was recorded in *Piptoporus* betulinus (Buxton 1960), usually develops on the bottom of a streamlet and the above-mentioned record of its larvae in fungi is not typical. Only two species of Achyrolimonia – A. neonebulosa (Alexander, 1924) and A. decemmaculata (Loew, 1873) are associated with fungi (Lindner 1958; Buxton 1960). A. decemmaculata was found directly in fungi, A. neonebulosa develops in mouldy wood. Atypophthalmus inustus

(Meigen, 1818) and *Rhipidia uniseriata* Schiner, 1864 only once were recorded developing in fungi (Stubbs & Chandler 1978) as they usually occur in rotten wood. Three species of *Discobola* are associated with fungi: Discobola annulata (Linnaeus, 1758) – a European species developing in mouldy wood and sometimes in fungi (Krivosheina et al. 1986); the Palearctic species – Discobola parvispinula (Alexander, 1947) was recorded in *Clavicorona pyxidata* once (Ševčik 2006); Discobola margarita Alexander, 1924 is distributed in southeastern Palaearctic and Oriental regions and develops only in mouldy wood (Krivosheina et al. 1986). Larvae of Limonia usually develop under wet leaf litter, but the species Limonia albifrons (Meigen, 1818), L. flavipes (Fabricius, 1787), L. nubeculosa Meigen, 1804, L. phragmitidis (Schrank, 1781) and L. trivittata (Schummel, 1829) were also found in mushrooms (Savchenko 1981; Lindner 1958; Krivosheina et al. 1986; Buxton 1960; Hackman & Meinander 1979). It is known that larvae of all Metalimnobia and Ula species usually develop in fungi (Lindner 1958; Buxton 1960; Hackman & Meinander 1979; Krivosheina et al. 1986). Larvae of Neolimonia dumetorum (Meigen, 1804) and Rhipidia maculata Meigen, 1818 develop in mouldy wood (Krivosheina et al. 1986), but there are a few records of their development in mushrooms as well (Dely-Draskovits 1972; Trifourkis 1977).

Larvae of crane flies developing in fungi were not purposively investigated in Lithuania earlier.

Five species of winter flies, *Trichocera annulata* Meigen, 1818, *Tr. hiemalis* (De Geer, 1760), *Tr. regelationis* (Linnaeus, 1758), *Tr. rufescens* Edwards, 1921 and *Tr. saltator* (Harris, 1776) were reported to develop in

fungi (Yakovlev 1994). Larvae of these species usually develop in rotting organic matter. *Tr. annulata* and *Tr. saltator* can develop in corpses (Krivosheina *et al.* 1986). It was proved that *Tr. annulata* feeds on muscles. However, in Lithuania larvae of this family have not been studied at all.

MATERIAL AND METHODS

Mushrooms with larvae were collected by S. Pakalniškis (SP), P. Ivinskis (PI), J. Rimšaitė (JR), R. Sprangauskaitė (RS) and V. Stukonis (VS) in 21 administrative districts and 41 localities in May-September 1997-2008. Alytus d., env. of Vidzgiris (N 54°22'44.8" E 24°00'06.6"); Akmenė d., env. of Peiliškės (N 56°15'30.7" E 22°37'31.6"); Anykščiai d., env. of Troškūnai (N 55°34'13.6" E 24°49'55.0"); Biržai d., env. of Spalviškiai (N 56°17'19.4" E 24°56'07.5"); Joniškis d., env. of Juodeikiai (N 56°13'55.9" E 23°12'25.7"); Kaunas d., env. of Pavejuonis (N 54°59'46.5" E 23°43'46.6"), env. of Ringovė (N 55°02'59.6" E 23°31'18.1"); Kaišiadorys d., env. of Strošiūnai (N 54°48'08.6" E 24°31'23.4"); Kėdainiai d., env. of Labūnava (N 55°09'02,9" E 23°59'00.7"); Kretinga d., env. of Kalno Grikštai (N 55°51'26,9" E 21°23'59.2"); Lazdijai d., env. of Gerdašiai (N 53°56'43.6" E 23°52'50.2"); Marijampolė d., env. of Opšrūtai (N 54°36'00.5" E 23°42'17.8"), Bukta forest (N 54°26'26.0" E 23°28'19.2"); Molètai d., env. of Šilai (N 55°03'31.7" E 25°33'31.6"); Plungė d., env. of Mikytai (N 56°07'26.6" E 21°24'32.7"); Plungė d., env. of Plateliai (N 56°07'45.5" E 21°59'45.2"); Prienai d., env. of Stanuliškės (N 54°33'09.2" E 24°24'43.2"); Radviliškis d., env. of Burūnai (N 55°31'39.3" E 23°34'07.2"); Šiauliai d., env. of Vilkuriai (N 55°51'38.6"E 23°13'19.6"); Švenčionys d., env. of Obelų Ragas (N 55°17'16.5" E 26°02'48.3"), env. of Pažeimenė (N 55°01'34.4" E 25°52'09.5"), env. of Purvynas (N 55°01'47.04" E 25°37'56.3"), env. of Žvyriai (N 55°18'51.6" E 26°30'01.3"); Tauragė d., env. of Dabrupinė (N 55°17'53.5" E 22° 06'43.8"); Trakai d., env. of Čižiūnai (N 54°35'43.1" E 24°33"54.2"), env. of Velnio Duobė (N 54°36'41.9" E 24°30'55.1"), env. of Varliškės (N 54°46'39.8" E 24°52'02.6"); Ukmergė d., env. of Vaisgėliškis (N 55°17'22.9" E 24°51'28.8"); Varena d., Čepkeliai forest (N 54°01'45.3" E 24°24'24.7"), env. of Darželiai (N 54°01'30.1" E 24°19'27.6"), env. of Dubininkas (N 54°06'03.1" E 24°16'36.8"), Puvočiai (N 54°06'45.0" E 24°18'12.7"); Vilkaviškis d., env. of Dabrovolė (N 54°28'46.0" E 22°46'48.7"), Vištytgiris forest (N 54°26'31.4" E 22°48'12.9"); Vilnius, Antavilis forest (N 54°47'01.9" E 25°25'06.1"), env. of Pilaitė (N 54°41'51.1" E 25°11'35.6"), env. of Smėlynė

(N 54°45'04.7" E 25°22'24.1"), env. of Visoriai (N 54°45'07.3" E 25°15'26.6"); Vilnius d., env. of Vaigeliškės (N 54°48'02.5" E 24°59'58.6"), env. of Dūkštos (N 54°49'36.2"E 24°57'27.8").

Methods of collecting and rearing larvae were described by Rimšaitė (1998, 2000). Adult crane flies were studied under a Zeiss SV dissecting microscope. The terminology of morphological features used in this paper generally follows that of Oosterbroek and Theowald (1991). The distribution area of crane flies given in the current article is in accordance with Oosterbroek (2010). The distribution of crane flies and winter flies in Lithuania is given according to Podenas (unpubl.). Habitats of crane flies larvae are presented by Podėnienė (unpubl.). A total of 500 adults of crane flies and 25 adults of winter flies were reared from mushrooms during these investigations. Adults were identified by S. Podėnas. Fungi were identified by J. Rimšaitė with the assistance of habil. Dr V. Urbonas, Dr E. Kutorga in accordance with 'Fungi of Lithuania' (Gricius & Matelis 1996; Urbonas 1997a, b, 1999, 2001; Kutorga 2000) and other atlases of fungi (Mazelaitis & Urbonas 1980; Bon 1988; Sterry 1995).

Each record is given in the following order: the species name of an insect – host fungus: the number of males, number of females, locality, sampling date/emergence date and collector's name.

RESULTS

Eight species of crane flies (*Limonia nubeculosa*, *Metalimnobia bifasciata*, *M. quadrimaculata*, *M. quadrinotata*, *Ula bolitophila*, *U. mollissima* Haliday, 1833, *U. mixta* Stary, 1983 and *U. sylvatica* (Meigen, 1818)) and two species of winter flies (*Trichocera hiemalis* and *Tr. rufescens*) were reared from fungi during the present investigation. Specimens of *U. mixta* were found in Lithuania for the first time. Trophic relationships of this species with fungi were revealed for the first time.

Family Limoniidae

Limonia nubeculosa Meigen, 1804

The species is common throughout Lithuania and was found in a variety of habitats. The flight period lasts from late April until early October. *L. nubeculosa* is widespread throughout the Holarctic region.

Adults were reared from Tricholoma sp.: $1 \circlearrowleft$, $1 \circlearrowleft$, environs of Mikytai, 27 July – September 2007, JR. **Habitats**: coniferous forests.

The fungi in which larvae were recorded are as follows: *Phlebia merismoides*, *Paxillus involutus*,

Lepista nebularis, *Flammulina velutipes* (Buxton 1960; Trifourkis 1977).

Metalimnobia bifasciata (Schrank, 1781)

This species is common throughout Lithuania and was found in a variety of habitats. The flight period lasts from May until September. *M. bifasciata* occurs in Palaearctic and Oriental regions. According to our data, it is the commonest species of crane flies developing in fungi in Lithuania.

Adults were reared from *Amanita rubescens*: 1, Šilai env., 18 July – 10 August 2002, JR; from Cortinarius sp.: 196, 25 and two specimens, Pavejuonis env., 27 July – 14 August 2000, JR; 3♂, 3♀ and 15 specimens, Vidzgiris env., 12 August – September 2000, JR; 2♀, Plateliai, 27 August – September – October 2007, JR; from *C. armillatus*: 3♂, 1♀ Čepkeliai env., 17 August–September 2000, JR; from Lactarius sp.: 3♂, 4♀, Kalno Grikštai env., 20 July – August 1999, JR; from L. deterrimus: 93, 5 Juodeikiai env., 7 July – 3 August 1998, JR; 1 \circlearrowleft , Labūnava env., 4–30 August 2000, JR; 34 specimens, Dabrupinė env., 2 August – 20 August – September 2000, JR; 9♂, 6♀, Peiliškės env., 4 September – October 2008, JR; from L. necator: 32, Dubininkas env., 1–27 July 1998, RS; 3♂, 8♀, Opšrūtai env., 9–20 August 2000, JR; $2 \circlearrowleft$, $7 \circlearrowleft$ and one specimen, Varliškės env., 27 September – November 2000, JR; from L. piperatus: 16, Spalviškiai env., 30 June -24 July 1998, JR; from L. scrobiculatus: 46, 19, Dabrupinė env., 2–23 August 2000, JR; from L. torminosus: 13, 39, Dūkštos env., 19 September – October 2003, JR; from *Leccinum scabrum*: three specimens, Labūnava env., 4 August – September 2000, JR; from *Polyporus squamosus*: $1 \stackrel{\frown}{\downarrow}$, $2 \stackrel{\frown}{\circlearrowleft}$, env. of Verkiai, 20–27 May – 5 June 2008, JR; from Russula delica: 3, Dūkštos env., 29 July – 25–31 August 2003, JR; from R. foetens: 16, Šilai env., 18 July – 22 August 2002, 1♂ 1♀, 18 July – 10 August 2002, JR; from Russula sp.: 1, Čižiūnai env., 23 June – 24 July 1998, SP; $4 \circlearrowleft$, $2 \circlearrowleft$, Žvyriai, 8 July – August–September 1999, JR; $5 \circlearrowleft$, $4 \circlearrowleft$, Dabrovolė, 11 July – 1 August 2002, JR; 1, Žvyriai env., 8 July – August 1999, JR; 53, 29, Bukta forest, 11 August – 25 August – September 2000, JR; 1♂, 1♀, Peiliškės env., 4 September – October 2008, JR; from Megacollybia platyphylla: 63, 1♀ and one specimen, Vaisgėliškis env., 19 July – 14–24 August 2000, JR; from *Paxillus involutus*: 1, Juodeikiai env., 7–31 July 1998, JR; 1♂, 1♀ from Piptoporus betulinus: Spalviškiai env., 28 May – 11-16 June 1998, VS; from Polyporus umbellatus: 5♀ and one specimen, Pavejuonis env., 27 July – August - September 2000, JR; from Suillus sp.: 2♀, Troškūnai env., 15 August – 22–26 September 2000, JR; from *Agaricales* s. l.: two specimens, Vilkuriai env., 4 August – 3 September 1999, JR; 1♂ Dūkštos env., 8 June – 8 July 1998, JR; 3♂, 2♀, Opšrūtai env., 9 August – September 2000, JR.

Habitats: fungi with larvae were collected in successive forest habitats: broadleaved deciduous forests (24% of samples with larvae), spruce forests (20%), oak-hornbeam forests (20%), swamp and bog woodland (16%), pine forests (12%) and mixed forests (8%).

Mushrooms in which larvae were recorded include: Morchella esculenta, Gyromitra infula, Scutiger ovinus, Phlebia merismoides, Inonotus radiatus, Trametes versicolor, Pseudotrametes gibbosa, Ganoderma applanatum, Aurantiaporus fissilis, Polyporus squamosus, Scleroderma cepa, Cantarellus cibarius, Lentinus lepideus, Suillus luteus, Xerocomus subtomentosus, Boletus pinicola, Leccinum versipelle, L. vulpinum, L. scabrum, Tricholomopsis platyphylla, Russula delica, Lactarius necator, L. resimus, L. terminosus, L. vellereus, Boletus edulis, Lyophyllum loricatum, Russula nigricans (Buxton 1960; Dely-Draskovits 1972; Trifourkis 1977; Hackman & Meinander 1979; Ševčik 2001, 2004; Yakovlev 1994).

Metalimnobia quadrinotata (Meigen, 1818)

This species is common in Lithuania and was found in a variety of habitats. The flight period lasts from late April until September. *M. quadrinotata* occurs in the Palaearctic region.

Adults were reared from *Helvella crispa*: 1♂ Burūnai 3 July – 20–30 July 2000, JR.

Habitats: fungi with larvae were collected in spruce forest.

Fungi in which larvae were recorded are as follows: *Gyromitra infula*, *Peziza badia*, *Clavulina cinerea*, *Amanita* sp., *Russula* sp. (Lindner 1958; Buxton 1960; Hackman & Meinander 1979; Yakovlev 1994).

Metalimnobia quadrimaculata (Linnaeus, 1761)

This species is common in Lithuania and was found in a variety of habitats. The flight period lasts from mid May until early October. *M. quadrimaculata* is known from the Holarctic region.

Adults were reared from *Pleurotus* sp.: 2 \circlearrowleft , Purvynas env., 27 September – October 2007, JR; from *Polyporus squamosus*: 1 \circlearrowleft , Verkiai env., 20–27 May – 5 June 2008, JR.

Habitats: fungi with larvae were collected in deciduous swamp forest and mixed forest.

Fungi in which larvae were recorded include: Gyromitra infula, Fomes fomentarius, Thelephora terrestris, Pseudotrametes gibbosa, Phaeolus schweinitzi, Laetipo-

rus sulphureus, Bjerkandera adusta, Abortiporus biennis (Coe 1941; Lindner 1958; Krivosheina et al. 1986; Yakovlev 1994; Ševčik 2001, 2006).

Family Pediciidae

Ula bolitophila Loew, 1869

This species is common in Lithuania. It was found in very different forests. The flight period lasts from May until August. *U. bolitophila* is known from the whole of the Palaearctic region.

Adults were reared from *Megacollybia platyphylla*: 1♀, Vaisgėliškis env., 19 July – 14 August 2000, JR; from *Piptoporus betulinus*: 4♂, 1♀, Spalviškiai env., 28 May – 11 June 1998, VS; from *Russula* sp.: 1♀, Juodeikiai env., 7 July – 3 August 1998, JR.

Habitats: fungi with larvae were collected in habitats of successive forests: mixed, broad-leaved deciduous and spruce forests.

Fungi in which larvae were recorded are as follows: Ganoderma sp., Trametes sp., Fomes sp., Pleurotus sp., Trametes hirsuta, Piptoporus betulinus, Hericum coralloides, Collybia sp., Laetiporus sulphureus, Peziza micropus, Bjerkandera adusta, Polyporus brumalis, P. badius, Pleurotus pulmonarius, Climacocystis borealis, Postia caesia (Krivosheina et al. 1986; Yakovlev 1994; Ševčik 2001, 2006).

Ula mixta Stary, 1983

Until now this species was not known from Lithuania. *U. mixta* is known just from the West Palaearctic (Czech Republic, Finland (south), Germany, Great Britain, Norway, Romania, Slovakia and Switzerland). The existence of a trophic relationship between larvae of this species and fungi was revealed for the first time.

Adults were reared from *Lentinus lepideus*: $7 \, \circlearrowleft$, $11 \, \circlearrowleft$, two specimens, Smėlynė env., 14 September 1997, JR; from *Ramaria flava*: $5 \, \circlearrowleft$, $6 \, \circlearrowleft$, four specimens, Ringovė, 27 July – August – September 2000, JR.

Habitats: fungi with larvae were collected in mixed forest.

Ula mollissima Haliday, 1833

This species is very rare in Lithuania (known from just three localities). It was found in deciduous forests, carst caves and forest meadows only. The flight period lasts from May until October. *U. mollissima* is widespread in the West Palaearctic.

Adults were reared from *Armillaria mellea*: 2♂, Vaigeliškės, 7 September – 19 October 1999, JR; from *Lactarius* sp.: 1♂, Kalno Grikštai env., 20 July 1999, JR; from *L. detterimus*: 1♀, Dabrupinė, 2 July – July – September 2000, JR; from *Paxillus involutus*: 4♂, 3♀,

Varliškės, 27 September – October 2000, JR; from *Polyporus squamosus*: 10♂, 11♀, Verkiai env., 20–28 May 2008, JR; from *Russula foetens*: 1♂, 1♀, Vidzgiris env., 18 July – 1–7 September 2002, JR; from *Russula* sp.: 6♂, 10♀, Vidzgiris, 12 August – September – October 2000, JR; 1♂, Peiliškės env., 4 September – October 2008, JR.

Habitats: fungi with larvae were collected in mixed and deciduous forests.

Mushrooms in which larvae were found include: Bjerkandera adusta, Hydnum erinaceum, Russula nigricans, Inonotus hispidus, Meripilus giganteus, Leucoporus sp., Polyporus squamosus, Pholliota alnicola, Ph. squarrosa, Collybia fuscipes, Lyophyllum decastes, Amanita excelsa, A. aspera, A. rubescens, Armillaria mellea, Oudemansiella mucida, Scleroderma sp., Pyptoporus betulinus, Hydnum repandum, Boletus pulverulentus, Leccinium verfuscipes, Lyophyllum decastes, Hypholoma fasciculare, Russula adusta, Tricholoma inamomeum, Polyporus badius, P. squamosus, Albatrellus ovinus, Pleurocybella porrigens, Bondarzewia montana, Megacollybia platyphylla, Grifola frondosa, Pleurotus pulmonarius, P. squamosus, Hydnum repandum, Lactarius scrobiculatus (Riel 1921; Lindner 1958; Trifourkis 1977; Stubbs & Chandler 1978; Ševčik 2001, 2006).

Ula sylvatica (Meigen, 1818)

This species is common in Lithuania. It was found in deciduous, mixed and spruce forests, shrubs, lakeshores and river banks, farmsteads. This species has two generations a year (from late spring to early summer and from late summer to early autumn). *U. sylvatica* is widespread all over the Holarctic region.

Adults were reared from *Amanita muscaria*: 200, 39, Antavilis forest, 7 September – 2–8 October 1998, JR; 20° , 39° , Obely Ragas, 26 September – October 1997, PI, SP; from *Cortinarius armillatus*: 1♀, Čepkeliai, 17 August – 21 September 2000, JR; from Gyrodon lividus: 12∂, 9♀, Staniuliškės env., 4 September – October 1997, JR; from *Hygrophorus* sp.: 1♀, Obelų Ragas env., 26 September - October - November 1997, PI, SP; from *Hypholoma fasciculare*: 20° , 19° , Gerdašiai env., 11 July – 8 August 2000, JR; from Lactarius torminosus: 2♂, 5♀, Čepkeliai, 17 August – 12 September 2000, JR; 11♂, 9♀ Darželiai env., 28 September – October – November 1997, SP; from *L. necator*: $3 \circlearrowleft$, $3 \updownarrow$ Pajuodžiai env., 12 September – October 1997, SP; 5♂, 8♀, Varliškės, 27 September – November 2000, JR; from L. detterimus: $2 \circlearrowleft$, $3 \circlearrowleft$ and one specimen, Strošiūnai env., 9 August – September – October 2000, JR; from *Megacollybia platyphylla*: 4\(\delta\), Velnio Duobė, 8 June – 7–15 July 2002, JR; from *Paxillus involutus*:

Varliškės env., 27 September – 1 November 2000, JR; from Russula sp.: 16♂, 8♀ Juodeikiai, 7–30 July – 3 August 1998, JR; one specimen, Čižiūnai env., 23 June – 24 July 1998, RS; 5♂, 2♀, Gerdašiai env., 11 August – 3 September 1999, JR; from *R. emetica*: 3♀ and two specimens, Darželiai env., October 1997 – 28 September 1998, RS; from Rozites caperatus: 2♂, 2♀, Troškūnai env., 15 August – October 2000, JR; 2♀ from Suillus sp., Pažeimenė, 17 September – October 1997, JR; from *Stropharia aeruginosa*: 4\$\display\$, 3♀, Strošiūnai env., 10 August – 6 September 2000, JR; from Verpa bohemica: 20♂, 15♀ and one specimen, Visoriai env., 6 May – 1–2 June 1998, JR; from Agaricales s. 1.: 12, Vaigeliškės env., 3 September - October 2000, SP; 1♂, 2 ♀ Pilaitė env., 14 May -8 June 1997, JR.

Habitats: fungi with larvae were collected in pine (29.4%), mixed (52.9%), spruce (5.9%), deciduous (5.9%) and swamp (5.9%) forests.

Mushrooms in which larvae were recorded are as follows: Morchella elata, Ptychoverpa bohemica, Gvromitra esculenta, Peziza sp., Scutiger ovinus, Ramaria aurea, Cantharellus cibarius, Lentinus lepidens, Suillus bovinus, S. luteus, S. variegatus, Xerocomus subtomentosus, Boletus edulis, B. pinicola, Leccinium versipelle, L. vulpinum, L. holopus, L. scabrum, Paxillus involutus, Hygrophoropsis aurantiaca, Hygrophorus erubescens, Camarophillus pratensis, Laccaria bicolor, L. laccata, Tricholoma albobrunneum, T. flavobrunneum, T. flavovirens, T. focale, T. saponaceum, Armillaria borealis, A. gallica, Lyophillium connatum, L. decastes, Melanoleuca melarealis, M. melaleuca, Lyophillum connatum, Panellus serotinus, Tricholomopsis platyphylla, Amanita rubescens, Agaricus bitorquis, Coprinus atramentarius, Psathyrella candolleana, P. vernalis, Stropharia cyanea, Hypholoma capnoides, Pholiota heteroclita, Ph. lenta, Kuehneromyces mutabilis, K. vernalis, Inocybe lacera, Hebeloma mesopliaceum, Dermocybe cinnamomea, D. semisanquinea, Cortinarius collinitus, C. mucosus, C. trivialis, C. brunneus, Lactarius flexuosus, L. necator, L. resimus, L. torminosus, L. trivialis, L. rufus, L. scrobicularius, L. spinosus, Polyporus squamosus, Meripilus giganteus, Tyromyces fissilis, Heteroporus biennis, Pseudotrametes gibosa, Bjerkandera adusta, Auricularia mesenterica, Russula cyanoxantha, R. paludosa, Peziza micropus, Sarcodon imbricatus, Albatrellus ovinus, Hydnum repandum, Paxillus involutum, P. filamentosum, Lactarius deterrimus, Hygrophorus sp., Pleurotus pulmonarius, Polyporus squamosus, Postia caesia, Pseudohydnum gelatinosum, Trametes versicolor (Buxton 1960; Yakovlev 1994; Bryce 1957; Ševčik 2001, 2006).

Family Trichoceridae

Trichocera (Trichocera) hiemalis (De Geer, 1760)

This species is common in Lithuania. It was found in shrubs, meadows, gardens, peat bogs and various types of forests, parks and cities. This species has two flight periods a year (from late February till mid-May and from mid-September till late October). *Tr. hiemalis* is widespread all over the Palaearctic region.

Adults were reared from Agaricales s. l.: 1♂, 1♀ Vištytgiris env., 16 October – November 2002, JR.

Habitats: fungi with larvae were collected in deciduous forest.

Fungi in which larvae were found include: *Trametes versicolor*, *Meripilus giganteus*, *Clavulinopsis vermicularis*, *Pleurotus ostreatus*, *Sarcodon imbricatum*, *Scleroderma citrinum*, *Entoloma nidorosum* (Buxton 1960; Trifourkis 1977; Hackman & Meinander 1979; Ševčik 2001).

Trichocera (Trichocera) rufescens Edwards, 1921

This species is common in Lithuania. It was found in shrubs, meadows, gardens, various types of forests and cities. The flight period lasts from late September until mid-November. *Tr. hiemalis* is widespread all over the Palaearctic region and was introduced into the Holarctic region.

Adults were reared from *Tricholoma equestre*: $6 \circlearrowleft$, $5 \circlearrowleft$, Puvočiai env., 2-25 October -3 November 1999, JR; $1 \circlearrowleft$, $3 \hookrightarrow$ and four specimens, Dubininkas env., 1 October -11 November 1999, RS; from *Russula nigricans*: $3 \circlearrowleft$, $2 \hookrightarrow$ Puvočiai env., 2 October -5 November 1999, JR.

Habitats: fungi with larvae were collected in pine forest

Fungi in which larvae were detected are: *Hypholoma sublateritium, Lepiota aspera, Pluteus cervinus, Armillaria gallica, A. cepistipes* (Buxton 1960; Ševčik 2001, 2004).

DISCUSSION

Separate species of crane flies use over 150 species of mushrooms as a substrate for their larvae development (Yakovlev 1994). The investigation conducted in Lithuania earlier revealed that larvae of *Ula bolitophila*, *Achyrolimonia decemmaculata* develop in fungus fruit bodies (Podėnas 1995). However, the data available in that study on other species associated with fungi (*Metalimnobia bifasciata*, *M. quadrinotata* (Meigen, 1818) were insufficient: the study merely indicated that these species develop in various fungus species. *M. quadri-*

maculata was mentioned as a species developing under the bark of *Salix* sp. trees (Table 1).

During our study larvae of *Ula mixta* were reared from fungi for the first time. This species developed in *Lentinus lepideus* and in *Ramaria flava*. Another species – *Metalimnobia bifasciata* was reared for the first time from seven fungus species such as *Piptoporus betulinus*, *Lactarius deterrimus*, *L. scrobiculatus*, *Paxillus involutus*, *Megacollybia platyphylla*, *Dendropolyporus umbellatus* and *Cortinarius armillatus*. Larvae of *Ula bolitophila* were for the first time reared from *Megacollybia platyphylla*. *Ula bolitophila* prefers lignicolous fungi (Jakovlev 1994), but we reared it from *Russula* sp. also.

A new trophic substrate – fruit bodies of the fungus *Tricholoma equestre*, *Russula nigricans* was established for *Trichocera* (*Trichocera*) rufescens. All literature records (Ševčík 2006) as well as our original records are from soft agarics growing in autumn. The new trophic substrate – fruit bodies of the fungus *Pleurotus* sp., *Polyporus squamosus* was established for *Metalimnobia quadrimaculata*. According to literature, this limoniid species is possibly associated with *Bjerkandera adusta*, but it was also reared from other fungus species, as well as from decaying wood (Jakovlev 1994), probably due to the pupation of larvae therein after the fungus had decomposed (Ševčík 2006).

Most larvae develop in decaying fungus fruit bodies. A total of nine species of crane flies and two species of winter flies are known to develop in fungi in Lithuania (Table 1).

Key to crane fly larvae (Tipuloidea: Pediciidae, Limoniidae) developing in mushrooms

Larvae of crane flies which have a hemicephalic head capsule and only two spiracles on the last abdominal segment are unique among other nematoceran Diptera developing in fungi, because the latter have more than one pair of spiracles and a eucephalic head capsule. Winter flies can be distinguished from the rest fly species by four-spiracled larvae (one pair situated on the first thoracical segment and the other one on the terminal abdominal segment). Larvae of flies belonging to the families Anisopodidae, Psychodidae and Ptychopteridae are known to be similar, but they cannot be found in fungi in Lithuania.

1 (18). Larva with two posteriorly situated spiracles. Head capsule hemicephalic Limoniidae, Pediciidae 2 (5). Spiracular field with five well developed lobes with dark sclerites (Fig. 1) *Ula* (2)

3 (4). Dorsal sclerite oval-shaped, with straight inner

Table 1. Crane flies and winter flies developing in mushrooms in Lithuania.

Fly species	Fungus species
Achyrolimonia	Daedalea quercina*
decemmaculata	
Limonia nubecu-	Tricholoma sp.
losa	-
Metalimnobia	Amanita rubescens, Cortinarius sp.,
bifasciata	Lactarius sp., L. deterrimus, L. neca-
	tor, L. s. piperatus, L. scrobiculatus,
	L. torminosus, Leccinum scabrum,
	Russula sp., Megacollybia platyphylla,
	Paxillus involutus, Piptoporus betuli-
	nus, Polyporus squamosus, Polyporus
	umbellatus, Suillus sp., Agaricales s. 1.
Metalimnobia	Under bark of Salix sp.*, Pleurotus sp.,
quadrimaculata	Polyporus squamosus
Metalimnobia	Various mushrooms*, Helvella crispa
quadrinotata	
Ula bolitophila	Megacollybia platyphylla, Polyporus
	adustus*, P. hirsutus*, P. pinicola*,
	P. fomentarius*, Piptoporus betulinus,
	Russula sp.
Ula mollissima	Russula sp., Lactarius sp., Armillaria
	mellea, Paxillus involutus
Ula mixta	Lentinus lepideus, Ramaria flava
Ula sylvatica	Amanita muscaria, Cortinarius
	armillatus, Gyrodon lividus, Hygro-
	phorus sp., Hypholoma fasciculare,
	Lactarius detterimus, L. necator,
	L. torminosus, Megacollybia platy-
	phylla, Paxillus involutus, Rozites ca-
	peratus, Russula sp., Suillus sp., Verpa
	bohemica, Stropharia aeruginosa,
	Agaricales s. 1.
	Agaricus*, Boletus*, Paxillus*, Can-
	tarelus*
Trichocera hiemalis	Agaricales s. l.
Trichocera rufe- scens	Tricholoma equestre, Russula nigricans

^{*-} by Podėnas (1995)

margin (Fig. 1) U. bolitophila

- 4 (5). Dorsal sclerite wedge-shaped, with elongated inner margin (Fig. 11) *U. sylvatica*, *U. mollissima* 5 (2). Spiracular field with more or less reduced lobes, which are without sclerites or with pale sclerites
- 6 (11). Hypostomal bridge with 11 teeth (Fig. 2). Spiracular field without sclerites or with hairy area between spiracles (Fig. 4) *Metalimnobia* (6)
- 7 (8). Spiracular field with large hairy area situated between the spiracles (Fig. 3) *M. bifasciata*
- 8 (7). Spiracular field without any hairy area (Fig. 12).
- 9 (10). Central frontal spike of the head capsule is shorter than the lateral one (Fig. 13) *M. quadrinotata* 10 (9). Central frontal spike of the head capsule is longer than the lateral one *M. quadrimaculata*

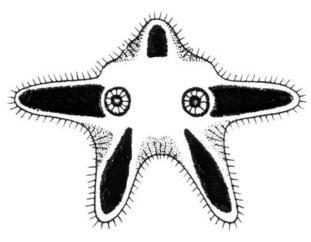


Figure 1. Spiracular field of *Ula bolitophila* (Savchenko 1986). Not to scale.

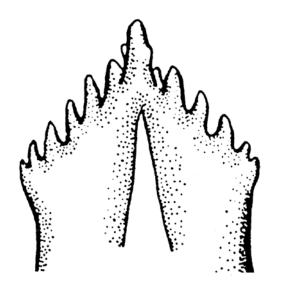


Figure 2. Hypostomal bridge of Metalimnobia quadrimaculata (Krivosheina & Mamaev 1967). Not to scale.

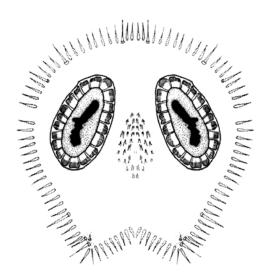


Figure 3. Spiracular field of *Metalimnobia bifasciata* (Cramer 1968). Not to scale.

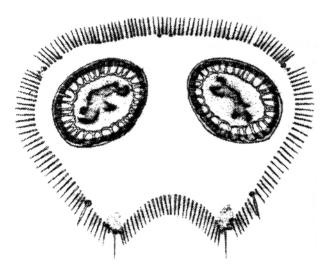


Figure 4. Spiracular field of Metalimnobia quadrinotata (Lindner 1958). Not to scale.

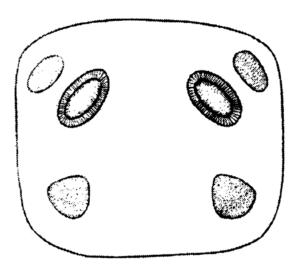


Figure 5. Spiracular field of Achyrolimonia decemmaculata (Savchenko 1985). Not to scale.

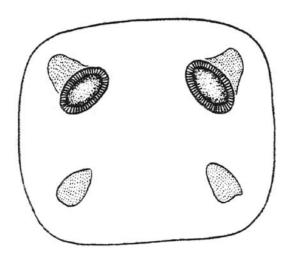


Figure 6. Spiracular field of *Limonia nubeculosa* (Brindle 1967). Not to scale.

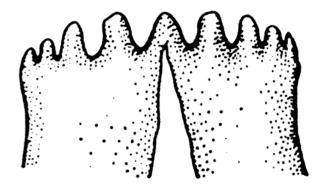


Figure 7. Hypostomal bridge of Discobola annulata (Krivosheina & Mamaev 1967). Not to scale.

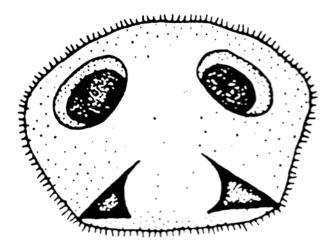


Figure 8. Spiracular field of *Discobola annulata* (Krivosheina & Mamaev 1967). Not to scale.



Figure 9. Hypostomal bridge of Rhipidia (Rhipidia) maculata (Reusch 1988). Not to scale.

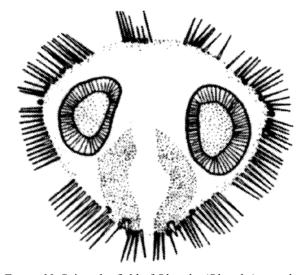


Figure 10. Spiracular field of Rhipidia (Rhipidia) maculata (Reusch 1988). Not to scale.

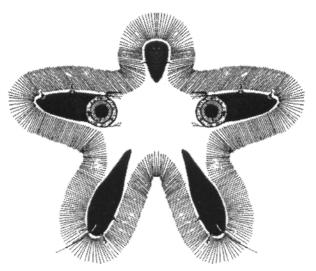


Figure 11. Spiracular field of *Ula mollissima* (Lindner 1959). Not to scale.

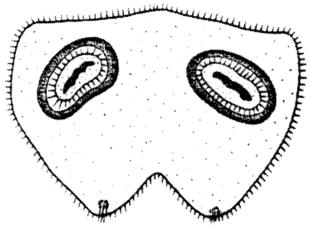


Figure 12. Spiracular field of Metalimnobia quadrimaculata (Savchenko 1985). Not to scale.

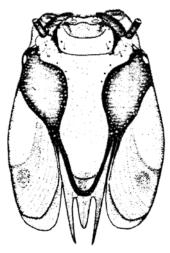


Figure 13. General view of the head capsule of *Metalimnobia quadrinotata* (dorsal view) (Lindner 1958).



Figure 14. General view of *Trichocera* sp. larva (lateral view) (Alexander 1981).

- 11 (6). Hypostomal bridge with 11 or fewer teeth. Spiracular field with two or four pale sclerites.
- 12 (15). Spiracular field with four pale sclerites.
- 13 (14). Dorsal sclerites are clearly separated from spiracles (Fig. 5) *Achyrolimonia decemmaculata*
- 14 (13). Dorsal sclerites are in contact with spiracles. *Limonia nubeculosa*
- 15 (12). Spiracular field with two pale sclerites.
- 16 (17). Hypostomal bridge with nine equal teeth, central tooth not protruded (Fig. 6). Spiracular field with two widely separated, wedge shaped sclerites (Fig. 7) *Discobola*
- 17 (16). Hypostomal bridge with 11 teeth (Fig. 8). Sclerites long, hooked and close to each other (Fig. 9) *Rhipidia*
- 18 (1). Larva with four spiracles (thoracical and posterior pairs). Head capsule eucephalic (Fig. 14) Trichoceridae.

REFERENCES

- Alexander, C. P. 1981. Trichoceridae. In: J. F. McAlpine, B. V. Peterson, G. E. Shewell, H. J. Teskey, J. R. Vockeroth and D. M. Wood (eds) *Manual of Nearctic Diptera* 1: 301–304. Ottawa, Ontario: Biosystematic Research Centre.
- Bon, M. 1988. *Pareys Buch der Pilze*. Hamburg, Berlin: Parey.

- Brindle, A. 1967. The larvae and pupae of British Cylindrotominae and Limoniinae (Diptera, Tipulidae). *Transactions of the Society for British Entomology* 17: 151–216.
- Bryce, D. 1957. Notes on the life histories of British Limoniidae (Diptera, Tipulidae). 2. *Ula sylvatica* Meigen. *Entomologist's Monthly Magazine* 93: 132–133.
- Buxton, P. A. 1960. British Diptera associated with fungi. 3. Flies of all families reared from about 150 species of fungi. *Entomologist's Monthly Magazine* 96: 61–94.
- Coe, R. L. 1941. Some breeding records of British Tipulidae (Diptera). *Entomologist's Monthly Magazine* 77: 173.
- Cramer, E. 1968. Die Tipuliden des Naturschutzparkes Hoher Vogelsberg. *Deutsche Entomologiche Zeitschrift* 15 (1–3): 133–232.
- Dely-Draskovits, A. 1972. Systematishe und ökologishe Untersuchungen an den in Ungarn als Schädlinge der Hutpilze auftretenden Fliegen. I. Limoniidae, Syrphidae, Platypezidae, Chloropidae (Diptera). *Acta Zoologica Academiae Scientarum Hungaricae* 18 (1/2): 7–21.
- Gricius, A. and Matelis, A. 1996. *Mushrooms of Lithuania*. *Aphyllophorales* 6 (2): 232 (in Lithuanian).
- Hackman, W. and Meinander, M. 1979. Diptera feedings as larvae on macrofungi in Finland. *Acta Zoologica Fennica* 16 (1): 50–83.
- Krivosheina, N. P. and Mamaev, B. M. 1967. Classification key to larvae of arboricolous dipteran insects. Moscow: Nauka. [Кривошенна, Н. П., Мамаев, Б. М. 1967. Определитель личинок двукрылых насекомых обитателей древесины. Москва: Наука.]
- Krivosheina, N. P., Zaicev, A. I. and Yakovlev, E. B. 1986. Insects damaging forest fungi in the European part of the USSR. Moscow: Nauka. [Кривошеина, Н. П., Зайцев, А. И., Яковлев, Е. Б. 1986. Насекомые – разрушители грибов в лесах Европейской части СССР. Москва: Наука.]
- Kutorga, E. 2000. *Mushrooms of Lithuania*. *Pezizales* 3 (5): 276 (in Lithuanian).
- Lindner, E. 1958. Pilzbewohnende Limoniiden Larven unter besonderer Berücksichtigung von *Limonia quadrinotata* Mg. (Diptera). *Tijdschrift voor Entomologie* 101: 263–281.
- Lindner, E. 1959. Beiträge zur Kenntnis der Larven der Limoniidae (Diptera). *Zeitschrift fur Morphologie und Okologie der Tiere* 48: 209–319.
- Mazelaitis, J. and Urbonas, V. 1980. *Mushrooms of Lithuania*. Vilnius: Mokslas. [Mazelaitis, J., Urbonas, V. 1980. *Lietuvos grybai*. Vilnius: Mokslas.]
- Oosterbreok, P. 2010. Catalogue of the crane flies of the world (Diptera, Tipuloidea: Pediciidae, Limoniidae, Cylindrotomidae, Tipulidae). URL http://ip30.eti.uva. nl/ccw/index.php.
- Oosterbroek, P. and Theowald, Br. 1991. Phylogeny of the Tipuloidea based on characters of larvae and pupae

- (Diptera, Nematocera). *Tijdschrift voor Entomologie* 134: 211–267.
- Podėnas, S. 1995. *The families Tipulidae, Limoniidae, Cylindrotomidae, Trichoceridae and Ptychopteridae in Lithuania: an eco-faunistic approach*. Thesis. Switzerland: University of Neuchatel.
- Reusch, H. 1988. *Untersuchungen zur Faunistik, Phänologie* und Morphologie der Limoniidae im Niedersächsischen Tiefland (Insecta, Diptera, Nematocera). Thesis. Germany: University of Hamburg.
- Riel, P. 1921. Les insectes parasites des champignons. I. Elevage et premiere liste des Dipteres fongicoles. *Annales Society Linnaeus Lyon* 67: 37–44.
- Rimšaitė, J. 1998. Initial data on fungus gnats (Diptera, Mycetophilidae) of Lithuania. *Acta Zoologica Lituanica* 8 (3): 60–62.
- Rimšaitė, J. 2000. Contribution to the knowledge of insects humificators of fungi in Lithuania. *Acta Zoologica Lituanica* 10 (1): 95–99.
- Savchenko, E. N. 1985. Limoniidae: subfamily Limoniinae. *The Ukraine Fauna* 14 (4): 1–180. Kiev: Naukova Dumka. [Савченко, Е. Н. 1985. *Комары-лимонииды.* Фауна Украины. Длиноусые двукрылые 14 (4): 1–180. Киев: Наукова Думка.]
- Savchenko, E. N. 1986. The Limoniid-flies (Introduction and subfamilies Pediciidae and Hexatominae). *The Ukraine Fauna* 14 (2): 1–340. Kiev: Naukova dumka. [Савченко, Е. Н. 1986. Комары-лимонииды (общая характеристика, подсемейства, педциины и гексатомины). Фауна Украины. Длиноусые двукрылые 14 (2): 1–340. Киев: Наукова Думка.]
- Savchenko, E. N. and Krivolutskaya, G. O. 1976. Limoniid-flies (Diptera, Limoniidae) of the South Kuril and South Sakhalin. Kiev: Naukova dumka. [Савченко, Е. Н., Криволуцкая, Г. О. 1976. Комары-лимонииды (Diptera, Limoniidae) Южных Курил и Южного Сахалина. Киев: Наукова Думка.]
- Stannius, H. 1829. Zur Verwandlungsgeschichte der Limnobia xanthoptera Meigen. *Beiträge Entomologishe Schles*. *Fauna* 1: 202–206.
- Sterry, P. 1995. *A Photographic Guide to Mushrooms of Britain and Europe*. London: New Holland Publishers.
- Stubbs, A. and Chandler, P. 1978 (eds). A dipterist's handbook. *The Amateur Entomologist* 15: 199–211.
- Ševčik, J. 2001. Diptera (excluding Mycetophilidae s. str.) associated with fungi in Czech and Slovak Republics: a survey of rearing records from 1998–2000. *Acta Universitatis Carolinae Biologica* 45: 157–168.
- Ševčik, J. 2004. New records of Diptera associated with fungi from Czech and Slovak republics. *Acta Facultatis ecologiae* 12: 135–142.
- Ševčík, J. 2006. Diptera associated with fungi in the Czech

- and Slovak Republics. Časopis Slezského Zemského muzea (A) 55 (2): 1–84.
- Trifourkis, S. 1977. *The bionomics and taxonomy of the larval Mycetophilidae and other fungicolous Diptera*. Ph. D. Thesis. London: University of London.
- Urbonas, V. 1997a. Mushrooms of Lithuania. Polyporales, Strobilomycetales, Boletales, Hygrophorales 8 (1): 200. Vilnius: Institute of Botany. [Urbonas, V. 1997. Lietuvos grybai. Kempiniečiai, žvynbaravikiečiai, baravykiečiai, guoteniečiai 8 (1): 200. Vilnius: Botanikos institutas.]
- Urbonas, V. 1997b. *Mushrooms of Lithuania. Tricholomatales* 8 (2): 216. Vilnius: Institute of Botany. [Urbonas, V. 1997. *Lietuvos grybai. Baltikiečiai* 8 (2): 216. Vilnius: Botanikos institutas.]
- Urbonas, V. 1999. *Mushrooms of Lithuania. Agaricales, Entolomatales* 8 (3): 296. Vilnius: Institute of Botany. [Urbonas, V. 1999. *Lietuvos grybai. Agarikiečiai, gijabudiečiai* 8 (3): 296. Vilnius: Botanikos institutas.]
- Urbonas, V. 2001. *Mushrooms of Lithuania. Amanitales, Russulales* 8 (4): 222. Vilnius: Institute of Botany. [Urbonas, V. 2001. *Lietuvos grybai*. Musmiriečiai, ūmėdiečiai. 8 (4): 222. Vilnius: Botanikos institutas.]
- Yakovlev, E. B. 1994. Palaearctic Diptera associated with fungi and myxomycetes. Petrozavodsk: Karelian research center Russian academy of sciences forest research institute. [Яковлев, Е. Б. 1994. Двукрылыые Палеарктики, связанные с грибами и миксомицетами. Петрозаводск.]

LIETUVOJE GRYBUOSE BESIVYSTANTYS ILGAKOJAI IR ŽIEMINIAI UODAI (DIPTERA: LIMONIIDAE, PEDICII-DAE, TRICHOCERIDAE)

V. Podėnienė, J. Rimšaitė, S. Podėnas

SANTRAUKA

Pateikiamas Lietuvos ilgakojų ir žieminių uodų (Limoniinae, Pediciidae, Trichoceridae), kurių lervos vystosi grybų vaisiakūniuose, sąrašas. Pirmą kartą sudarytas Lietuvos grybų, naudojamų kaip lervų vystymosi substratas, sąrašas, bei nustatyti *Metalimnobia bifasciata* (Schrank, 1781) trofiniai ryšiai su septynių rūšių grybais, o taip pat *Ula bolitophila* Loew – vienas naujas lervų mitybinis grybas. Tyrimų metu rasta nauja Lietuvos faunai rūšis – *Ula mixta* Stary, 1983 bei pirmą kartą nustatyti šios rūšies trofiniai ryšiai su grybais.

Received: 26 April 2010 Accepted: 9 September 2010